

Sleep and Oligodendrocyte Functions

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Abstract Transcriptomic studies have revealed that the brains of sleeping and awake animals differ significantly at the molecular level, with hundreds of brain transcripts changing their expression across behavioral states. However, it was unclear how sleep affects specific cell types, such as oligodendrocytes, which make myelin in the healthy brain and in response to injury. In this review, I summarize the recent findings showing that several genes expressed at higher levels during sleep are involved in the synthesis/maintenance of all membranes and of myelin in particular. In addition, I will discuss the effect of sleep and wake on oligodendrocyte precursor cells (OPCs), providing a working hypothesis on the function of REM sleep and acetylcholine in OPC proliferation.

Keywords Wake · REM sleep · Sleep deprivation · Bactrap · Myelin · Acetylcholine · Oligodendrocyte precursor cells · Neurogenesis · Multiple sclerosis

Introduction

Oligodendrocytes together with astrocytes constitute macroglia [1]. The main function of oligodendrocytes is to provide insulation and support for most of the axons in the CNS [2]. A single oligodendrocyte can enwrap around 20 axonal segments on average, providing multiple membrane lamellae in each segment [3]. The maintenance of vast amounts of plasma membrane is metabolically demanding,

and among all brain cells, oligodendrocytes are the ones showing the highest level of metabolic activity [4].

Myelin production is a key function of oligodendrocytes. Myelin is an insulation material constituted by 80 % lipid and 20 % protein that forms a layer, the myelin sheath, enwrapping the axon [5]. The main function of the myelin sheath is to protect and insulate the axon from the extracellular environment, thus allowing fast impulse propagation along the axon [6].

Sleep is a tightly regulated phenomenon, essential for all animal species carefully studied so far [7], and its loss leads to negative consequences for the brain and the entire body [8, 9]. Despite the importance of sleep, very little is known about the effect that sleep exerts on oligodendrocytes and myelin production. In the first part of this review, I will summarize the current knowledge on this topic, emphasizing the protective role of sleep on myelination. In the second part, I will provide some insights about the role of sleep on the proliferation of the oligodendrocyte precursor cells (OPCs).

Oligodendrocyte Functions are Modulated by Sleep and Wake

The first indication that some oligodendrocyte functions were regulated by sleep and wake came from gene expression studies, which showed that the expression of many glial genes was different during sleep and wake [10–17]. In 2004, Cirelli and collaborators found that sleep was associated with the transcription of several genes coding for myelin structural proteins (MOBP, MAG, plasmalogen, CD9), myelin-related receptors (insulin-like growth factor binding protein 2), and enzymes (2':3'-cyclic nucleotide-3'-phosphodiesterase, Na/K ATPase subunit α 2, methionine adenosyltransferase, carbonic anhydrase II). In addition, they observed a sleep associated increased transcription of genes coding for enzymes involved

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in the synthesis and transport of cholesterol, a major constituent of myelin and other membranes (thiolase, 3-hydroxy-3-methylglutaryl-Coenzyme A synthase, squalene synthase, lanosterol 14 α -demethylase) [11]. Few years later, Mongrain and colleagues also pointed out a similar association between sleep and genes involved in lipid metabolism and myelination. Importantly, to rule out the confounding role of the stress hormone corticosterone during sleep deprivation, they performed gene expression analysis also in adrenalectomized mice in which *opalin* and *plasmolipin*, both genes involved in myelination [18, 19], were found to be downregulated in sleep deprivation relative to sleep [16]. Another study, aimed to assess the transcriptomic profile of chronic sleep loss, demonstrated that long-term sleep deprivation downregulated the expression of two myelin-related genes, one coding for plasmolipin and the other for CD9, a membrane protein normally expressed in the mature myelin sheath, again suggesting a role for sleep in promoting myelination [13].

However, those gene expression studies had several limitations. First, the gene expression analysis was based on transcripts that were pooled from all brain cells, whereas in situ hybridization experiments have indicated that the effects of sleep and wake can vary significantly depending on cell type [20]. Another limitation was the exclusive assessment of mRNA levels, which are not always predictive of protein levels [21, 22]. To overcome these limitations, our group recently used the translating ribosome affinity purification (TRAP) technology and microarray analysis to obtain a genome-wide mRNA profiling of oligodendrocytes as a function of sleep, wake, and acute sleep deprivation [23]. In addition, since the TRAP method targets mRNAs already attached to ribosomes, a more functional (translational) view of cellular function could be achieved [24].

Gene expression analysis performed on oligodendrocyte-enriched samples of mouse forebrain showed that the expression of hundreds of transcripts ready to be translated into proteins changes because of sleep and wake in oligodendrocytes, and many wake-related and sleep-related genes have complementary or opposite functions. Specifically, many genes implicated in apoptosis, cellular stress and metabolism were upregulated during wake, whereas genes involved in phospholipids synthesis and myelination were transcribed preferentially during sleep [23], confirming the previous results on whole samples [11, 16]. Among these, both *opalin* and *plasmolipin* showed increased expression. In addition, we found *Qk*, a sleep-related gene coding for the RNA-binding protein Quaking that regulates several myelin-related transcripts at the posttranscriptional level [25, 26].

A link between sleep and myelin is consistent also with a recent hypothesis that assigns an energetic role to myelin [27]. Starting from the evidence that several components of the respiratory chains are expressed in myelin [28, 29], Morelli and colleagues proposed that the energy required for the

axonal function could be provided by the myelin sheath itself [30]. However, the intense oxidative phosphorylation in the myelin sheaths occurring during wake would lead to a local accumulation of protons. Therefore, during sleep, one of the supposed role of the myelin basic protein (MBP) and phospholipids forming the myelin would be to buffer the protons generated during wake [27, 30]. The general idea that the oligodendrocytes metabolically support the axons has been confirmed by several lines of evidence [31, 32]. However, recent data suggest that oligodendrocytes may support axons through mechanisms independent of myelination [33]. Reportedly, the dysmyelinated *shiverer* mutant mouse, which lacks the myelin basic protein, exhibited no obvious alteration of axonal integrity [34], suggesting that myelination and axonal support are two distinct oligodendrocyte functions. In addition, there is evidence demonstrating that lactate shuttling from the oligodendrocyte to the axon is a key mechanism through which axons are “fed” [33]. In this respect, particular attention has been given to the lactate transporter MCT1 that excretes lactate outside the cell [32] and is highly expressed in oligodendrocytes [35]. Interestingly, *Slc16a1*, the gene coding for MCT1, was found upregulated in sleep oligodendrocyte-enriched samples [23], suggesting that axon refueling might preferably occur in a condition of reduced axonal firing.

In conclusion, all together, these studies indicate that sleep may represent a favorable moment for membrane remodeling and myelination, and, conversely, sleep loss may negatively impact the oligodendrocyte physiology, thereby leading to cell dysfunctions that can undermine the axonal integrity.

OPCs Dynamics in Sleep and Wake

OPCs represent ~5 % of all cells in the brain [36]; they actively proliferate during development and, albeit at lower rate, during the adult life [36–38]. They mediate myelin formation in healthy brain and are rapidly recruited to prime the recovery process in response to injury [39]. Importantly, recent evidence showed that OPC proliferation is inhibited by extracellular glutamate through AMPA-mediated excitatory currents in OPCs, thus suggesting that neuronal activity can affect myelination and possibly drive other still unknown functions of OPCs [36–38].

Cortical neuronal activity [40], as well the expression of AMPA receptors [40], and their currents [41] increase during wake and decline during sleep. Using in vivo amperometry in the cerebral cortex, a parallel trend of extracellular glutamate levels was observed across the sleep/wake cycle [42], confirming that the glutamatergic tone is higher during wake than during sleep. These findings led us to test the hypothesis that OPCs dynamics could be affected by sleep and wake.

Gene expression analysis performed on oligodendrocyte-enriched samples of mouse forebrain showed that many genes

that changed their expression in sleep and wake were actually OPC specific genes. A further functional clusterization more focused on OCP genes revealed that genes promoting OPC proliferation were mainly upregulated during sleep [23]. Among these, *Yap1*, *Mxd3*, and *Nrg2* have been all linked to OPC proliferation [43–45]. By contrast, genes mostly involved in OPC differentiation and apoptosis were preferably transcribed during wake [23]. It is worth noting that *YY1* and *Sox17* are wake genes, and both implicated in OPC maturation, as it has been shown that interfering with their expression arrests the OPC differentiation towards more mature stages [46, 47].

Additional assessments based on immunocytochemical OPC characterization in combination with the BrdU assay, a specific method to evaluate cell proliferation, demonstrated that the number of newly formed OPCs increased during sleep relative to spontaneous wake and sleep deprivation. Finally, EEG studies in conjunction with BrdU assessments revealed a positive correlation between the level of OPC proliferation and the amount of REM sleep, indicating that this stage of sleep could represent a favorable condition for OPC proliferation [23].

Is REM Sleep Regenerative for OPCs?

The idea that sleep may promote cell proliferation is not new. Most of the studies have focused on understanding how sleep regulates hippocampal neurogenesis in adult mammals [48–57]. In a recent review, Mueller and colleagues concluded that much of the evidence supports the idea that sleep disruption has a negative impact on cell proliferation and, specifically, on neurogenesis [58]. The mechanisms by which this occurs are not clear, but there are indications that some “wake” factors, including corticosterone and interleukin 1 β , may mediate this effect [51, 59]. This would suggest that the presence of these factors is likely to be detrimental for neurogenesis and, eventually, for cell proliferation in general, weighing against the hypothesis that favoring neurogenesis is a function of sleep [58].

However, several studies showed that this might not be the case for REM sleep, which may have a more active role. A recent work performed using a selective REM sleep deprivation apparatus capable of producing an arousal stimulus as soon as some EEG features for REM sleep were met found a 63 % reduction in cell proliferation in the dentate gyrus [60]. The rate of proliferation also correlated with the amount of time spent by the mice in REM sleep [60]. In another study, 72 h of REM sleep deprivation led to a delayed rebound in neurogenesis 1 week after the deprivation; importantly, this increase was present also in adrenalectomized mice [51]. Since REM sleep rebound can persist for weeks after long-term REM deprivation [61], it is possible that the observed

increase in REM sleep had a positive modulation on neurogenesis, independent of corticosterone.

REM sleep is characterized by rapid and random movements of the eyes, muscle atonia, and high-frequency low-voltage EEG [62]. From a neurochemical point of view, REM sleep is characterized by low levels of neuromodulators with the exception of acetylcholine (ACh), whereas in NREM sleep, ACh is also low [63, 64]. Therefore, if some positive functions on cell proliferation are associated with REM sleep, ACh itself might carry them out.

Indeed, there are several lines of evidence indicating that ACh regulates neurogenesis in the hippocampus (see [65•] for a review). Immunotoxic ablation of basal forebrain cholinergic neurons projecting to the hippocampus strongly reduces the number of proliferating BrdU immunoreactive cells in the dentate gyrus [66]. Moreover, transgenic mice overexpressing a recombinantly inactivated form of synaptic ACh-esterase showed significantly reduced ACh hydrolysis and enhanced proliferation of progenitor cells, suggesting that ACh and ACh-esterase are important regulators of proliferative activity [67]. These results have been confirmed also in a recent experiment performed in vitro, in which a potent effect of ACh on neuroblast production has been discovered from postnatal sub-ventricular zone (SVZ), another region of the forebrain where neurogenesis occurs in addition to the dentate gyrus [68•]. Furthermore, the same study found a previously unknown population of choline acetyltransferase (ChAT) (+) neurons, which provide innervation of the SVZ and release ACh locally in an activity-dependent manner. Optogenetic inhibition and stimulation of these cells in vivo showed that they were capable of controlling neurogenic proliferation in SVZ [68].

SVZ is a critical region also for OPCs proliferation [36, 38, 69]. In addition, several in vitro studies demonstrated the ACh favors not only neurogenesis but also OPCs proliferation and survival [70, 71, 72•, 73, 74]. Exposure of primary OPC cultures to ACh led to an augmented OPC proliferation via the activation of ACh M1, M3, and M4 muscarinic receptors on OPCs [72•]. Since muscarinic receptors on OPCs are developmentally regulated, being highly expressed mainly in immature cells, ACh may contribute to the maintenance of an immature proliferating OPC pool in the brain [72]. Other findings, in addition to support the role of ACh as a growth regulatory signal for OPC proliferation, identified the molecular pathway that mediates this effect, which involves the protein kinase C activation and MAPK phosphorylation [70, 73].

In conclusion, these studies support the hypothesis that REM sleep may exert a positive modulation on OPC proliferation in forebrain, likely through the action of ACh (Fig. 1). However, many steps of this process need further testing. For example, the notion that ACh induces OPC proliferation is only supported by in vitro studies. In addition, it is still unclear

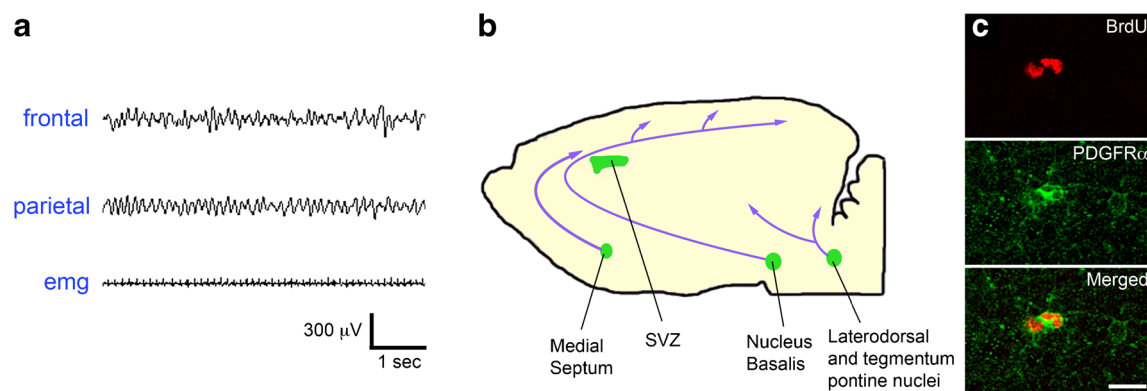


Fig. 1 Key elements of the hypothetical mechanism leading to OPC proliferation, which involves REM sleep and acetylcholine release. **a** Example of REM sleep as it appears in the mouse EEG. Note the intense theta activity in frontal and parietal derivations associated with muscle atonia (EMG, electromyogram). **b** Schematic section of a mouse brain, showing the locations of the most important groups of acetylcholine neurons and their projections to the forebrain.

why during wake, a state in which the Ach tone is elevated as much as REM sleep, OPC proliferation seems to be inhibited. In vitro experiments have indicated that the presence of noradrenaline, the other major wake neuromodulator, may induce cytotoxicity and even apoptosis in OPCs through oxidative stress [75]. If these results were confirmed in vivo, during wake, the positive action of Ach on OPC proliferation could not take place because of the counteracting action of noradrenaline.

Oligodendrocytes and OPCs Pathology and Sleep

If sleep plays a role in supporting some important oligodendrocyte functions, like myelination, and exerts a promoting role in the proliferation of new immature oligodendrocytes, then it is natural to ask whether sleep could be protective in diseases characterized by oligodendrocyte dysfunction. Unfortunately, there is not yet a definitive answer to this question. For instance, patients affected by obstructive sleep apnea show white matter alterations [76–78]. Although intermittent hypoxia has likely a major role in this case [79], sleep fragmentation associated with this disease can be also important and can negatively impact the oligodendrocyte physiology. Along the same line, a recent postmortem study revealed decreased myelination in patients suffering from restless leg syndrome [80], a disease often associated with insufficient sleep [81]. Although a direct demonstration that chronic sleep restriction can lead to oligodendrocyte dysfunction and myelin alteration is still lacking, these studies suggest that sleep loss may be detrimental to oligodendrocytes and more specifically to myelin. In multiple sclerosis (MS), an inflammatory autoimmune disease characterized by CNS demyelination, sleep disturbances are occurring in roughly 50 % of the

SubVentricular Zone (SVZ) was included among the cholinergic areas, because it displays locally cholinergic innervation. **c** Example of proliferating OPCs stained with the specific OPC marker PDGFR α (green) and positive to bromodeoxyuridine (BrdU, red), a synthetic nucleoside commonly used in the detection of proliferating cells in living tissues. Scale bar: 15 μ m

patients [82–85]. However, it is unknown whether sleep problems are only a consequence of the illness or if they actively contribute to the progress of the disease. Clinical studies have reported that sleep problems in MS are often under diagnosed [86] and may be related to daytime fatigue experienced by most of the patients [87]. There is also some evidence showing a positive correlation between the severity of the clinical picture and the presence of sleep problems [83], suggesting that the correction of sleep problems might help to improve the clinical conditions of MS patients. However, if this beneficial role of sleep is mediated by the mechanisms previously described on myelin and OPC proliferation remains unclear and needs further investigations.

Compliance with Ethics Guidelines

Conflict of Interest Michele Bellesi declares that he has no conflict of interest.

Human and Animal Rights and Informed Consent This article does not contain any studies with human or animal subjects performed by any of the authors.

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